

AN AQUATIC SYSTEM MULTI-LEVEL EVALUATION USING SPECIES-SPECIFIC SENSITIVITY BIOINDICATORS



Laura FEODOROV

University of Agronomic Sciences and Veterinary Medicine of Bucharest, Faculty of Biotechnology
Control Pollution Department, National Research and Development Institute for Industrial
Ecology—ECOIND

One Health

Student International Conference

Under the Patronage of the Romanian Academy
3-5th December 2025 | București, România



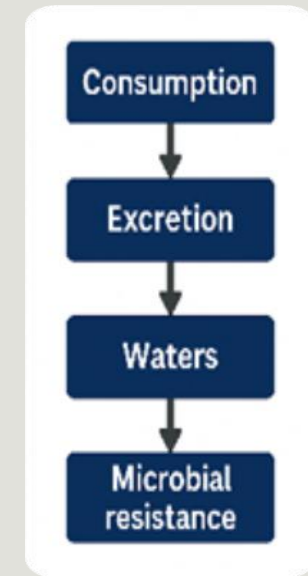
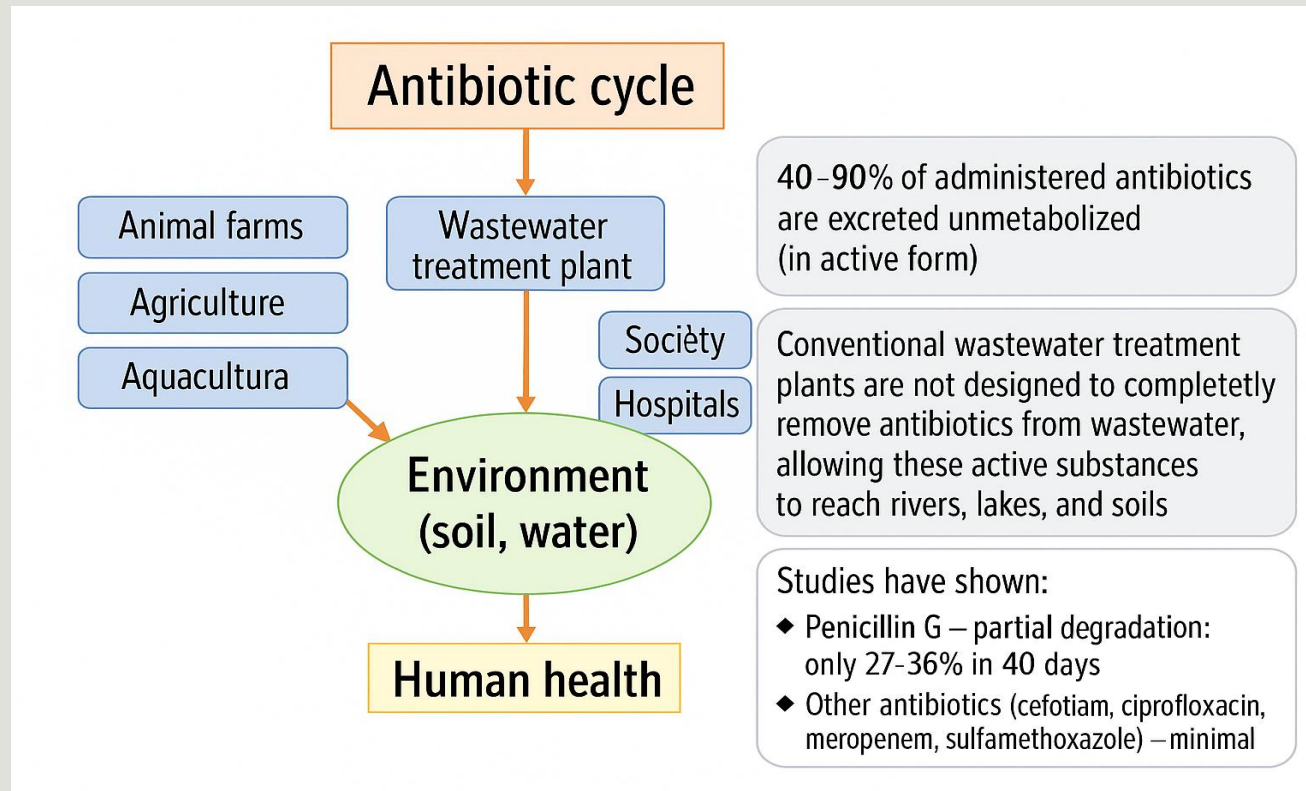
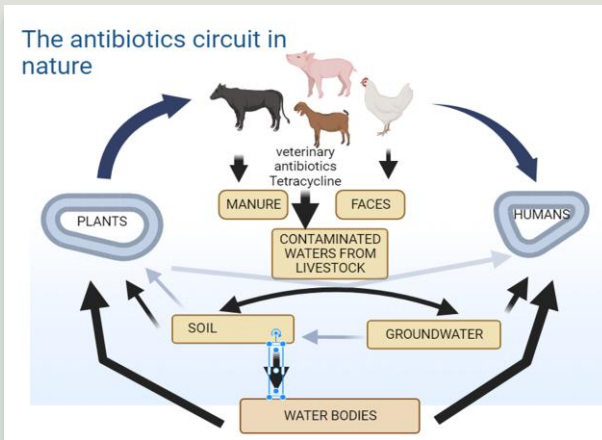
Antibiotic Pathways in the Environment

Medical use → Excretion → Wastewater → Natural waters

✓ Sources

✓ Entry routes

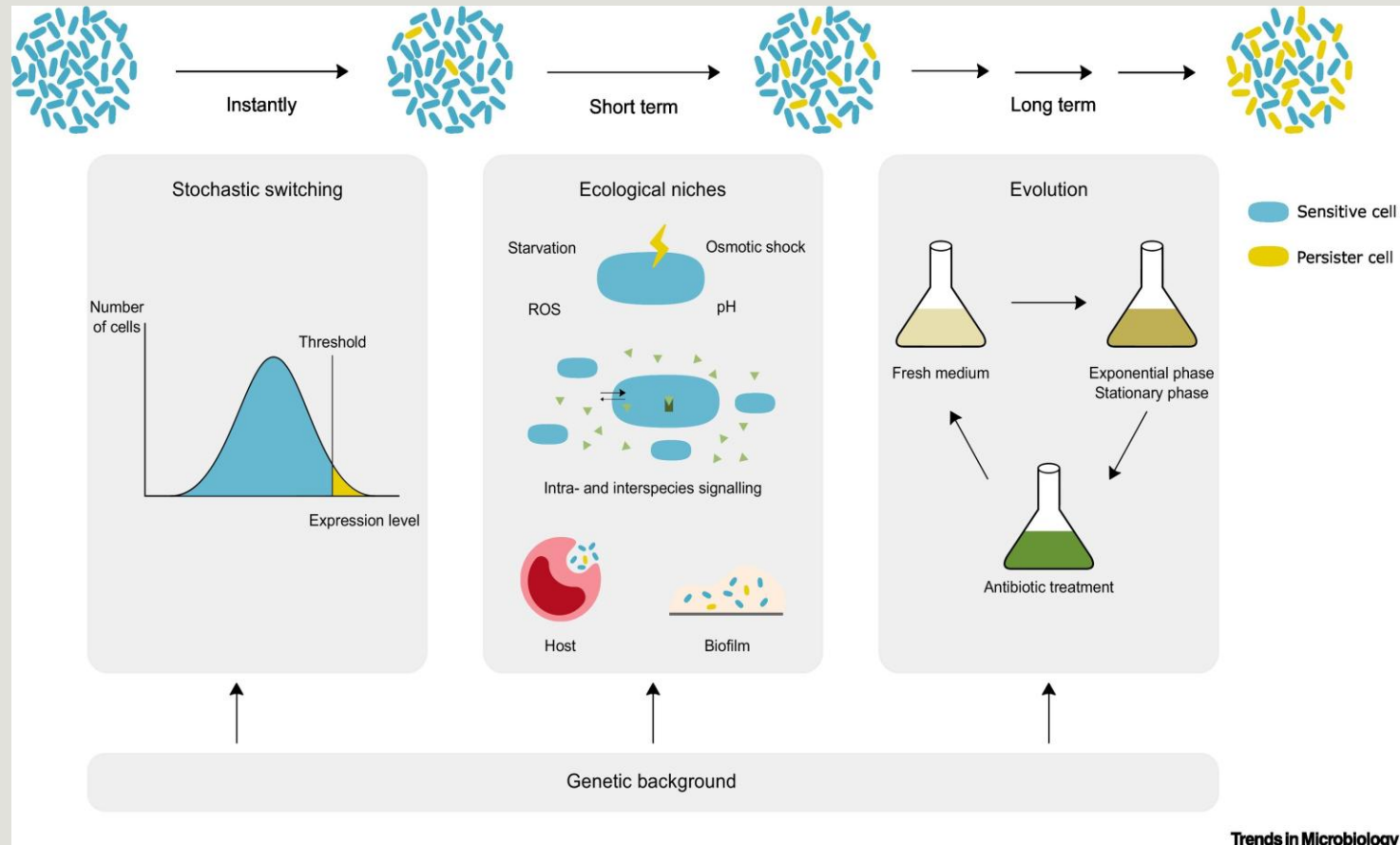
✓ Point vs non-point contamination



Environmental Fate of Antibiotics

Environmental persistence and microbial disturbance

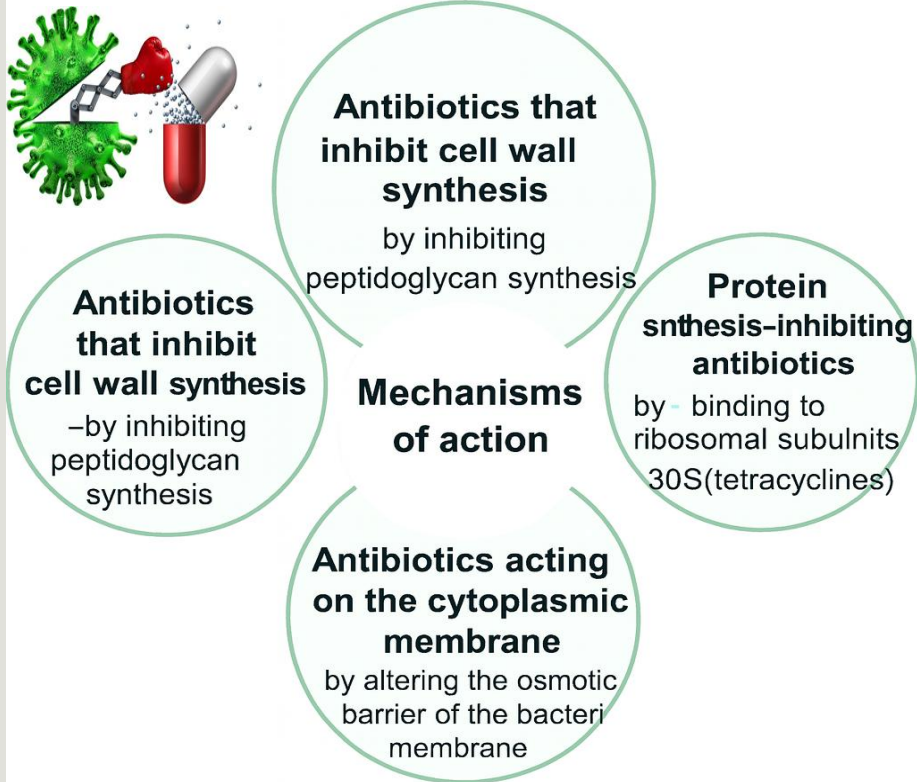
- ✓ Persistence
- ✓ Transformation
- ✓ Bioaccumulation



Ecological Implications of Antibiotic Pollution

- ✓ Microbial disruption
- ✓ Ecosystem imbalance
- ✓ AMR acceleration

Mechanism of action and effects of antibiotics on aquatic organisms



Effects of Antibiotics on Aquatic Organisms

Crustaceans

Acute test:

- Cefotaxime
– EC₅₀ = 25.582 mg/L
highly toxic
- Cefixime –
– EC₅₀ = 77.9.9 mg/L
moderate toxicity
- Ciprofloxacin –
– EC₅₀ ≈ 36.5 mg/L
oxidative effect and neurotoxicity

Chronic:

- Photosynthesis inhibition
- Oxidative stress
- Damage to pigments and membranes

Algae

- Antibiotics
 - oxytetracycline,
 - clarithromycin,
 - ciprofloxacin,
 - sulfamethoxazole

Fish

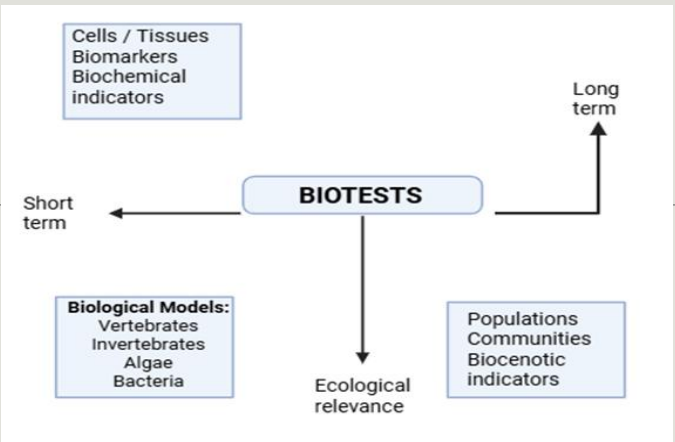
- Genetic, histological and enzymatic
- Enzymes hepatic – indicators of liver damage
- Quinolones - affect development and healing rate
- Sulfonamides – increase mortality disrupt microbiota and metabolism



Aquatic System Toxicity Evaluation

Tetracycline & Co-trimoxazole
Assessment

Daphnia magna & Vibrio fischeri



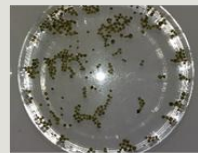
Fish



Daphnia



Algae



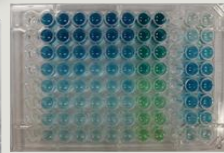
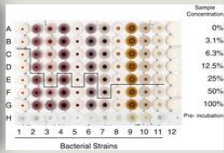
Aquatic plant



Earthworms



Higher plants



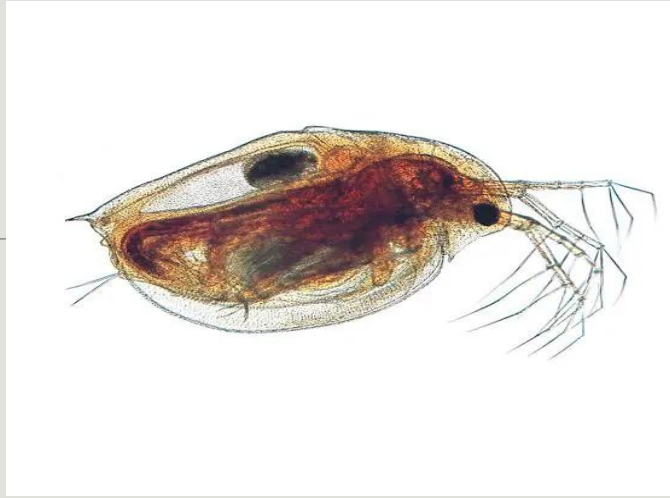
Bacteria

OECD/ ISO/EPA
Testing and interpretation Methodologies

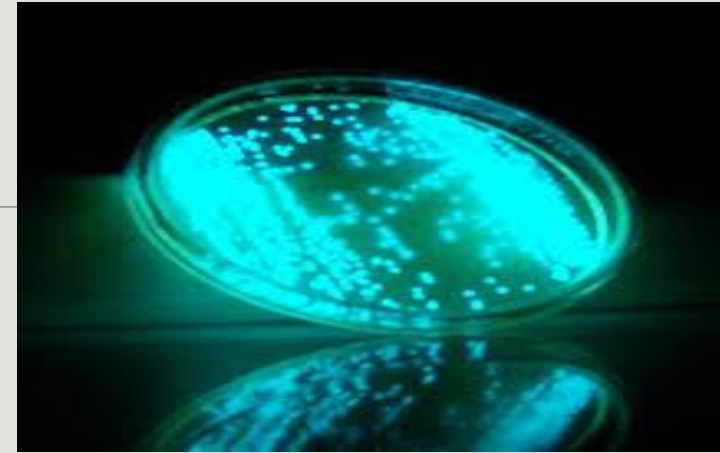
EC50, EC20, LOEC, NOEC, PEC, PNEC, risk coefficients, ecotoxicological limits based on the most detrimental case



Bioindicators used and study objectives



Daphnia magna:
sensitive crustacean



Vibrio fischeri:
bioluminescent bacterium used
in Microtox[®] assays



Assess acute and chronic toxic effects.



Determine EC50, NOEC and LOEC.



Evaluate adaptive responses to sublethal stress.



Materials & Methods

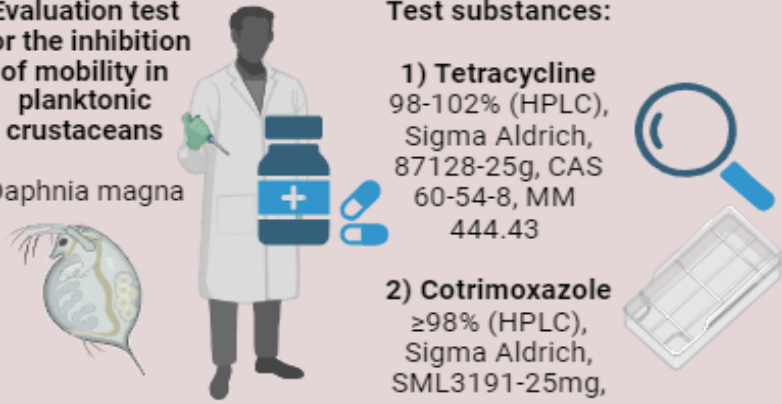
- Microtox® luminescence assay

Organism: <i>V.fischeri</i> (ATCC 7744)	Acute testes: 1 mg/L-1 gr/L) 30 min	Chronic testes: 48 h	Bioluminescence inhibition measured by Microtox® → EC ₅₀ via 4PL model
---	-------------------------------------	----------------------	---

- Daphnia acute and chronic exposure tests

Evaluation test for the inhibition of mobility in planktonic crustaceans

-Daphnia magna



Test substances:

1) Tetracycline
98-102% (HPLC), Sigma Aldrich, 87128-25g, CAS 60-54-8, MM 444.43

2) Cotrimoxazole
≥98% (HPLC), Sigma Aldrich, SML3191-25mg, CAS 8064-90-2, MM 1556.71

Method	Biological material	Test conditions	Tested concentrations	Monitored parameter	Final result
Acute toxicity					
OECD 202/ SR EN ISO 6341:2013/ Daphtoxkit F magna kit	<i>Daphnia magna</i> (water fleas) Age 90 h	Test duration 48h Incubation at 20±2°C, light Control with culture medium	Tetracycline 6.25-100mg/L Cotrimoxazole 6.25-100mg/L	Mortality/ immobilization	EC50-48h (mg/L) and <10% effect in control

- Statistical models: ANOVA, 4PL

- Concentrations: environmentally relevant ranges



Results

- *Vibrio fischeri*:

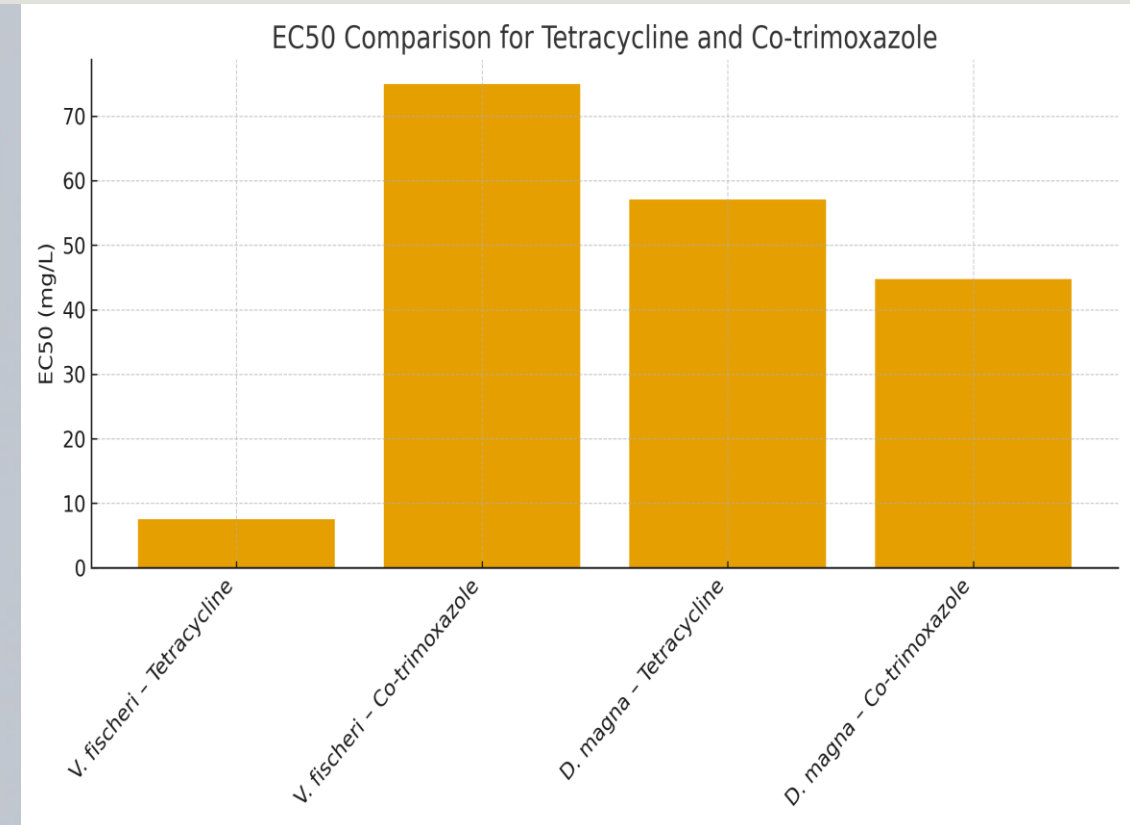
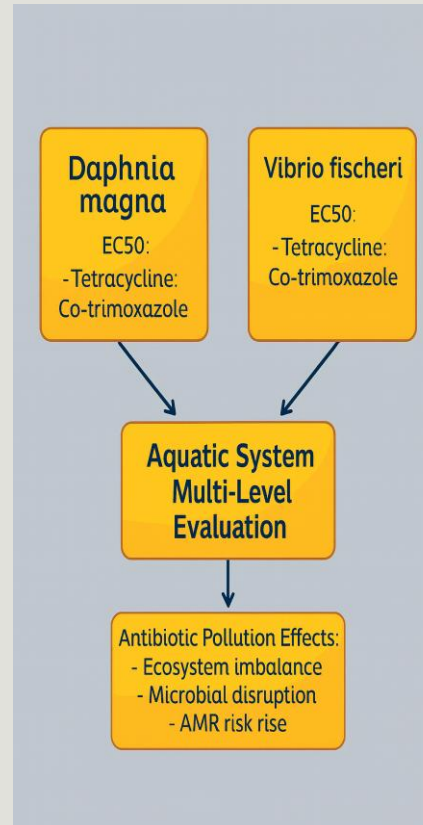
- Tetracycline EC50 = 5–10 mg/L
- Co-trimoxazole EC50 = 50–100 mg/L
- Significant inhibition of bioluminescence

- *Daphnia magna*:

- Tetracycline EC50 = 57.10 mg/L
- Co-trimoxazole EC50 = 44.76 mg/L
- Chronic exposure reduced mobility

- Comparative Insights:

- *Vibrio* detects rapid metabolic disruption
- *Daphnia* reveals ecological-level chronic impacts



Conclusions

Antibiotics pose complex ecological risks by disrupting microbial communities, impairing aquatic organisms, and promoting antimicrobial resistance.

Mitigations:

- Wastewater regulations must be strengthened.
- Advanced treatment technologies are essential.
- Reducing antimicrobial resistance requires integrated action.



Future Directions

- Further Toxicity Testing on Other Species
- Mechanistic Studies
- Chronic Toxicity Studies
- Mitigation Strategies
- Risk Assessment
- Regulatory Considerations
- Synergistic/ Antagonistic Effects



This study supports the **integration of biosensor-based tools** in environmental monitoring programs, promoting **early detection of antibiotic pollution** and contributing to **sustainable water management and ecosystem protection**.



THANKS

Călina-Petruța CORNEA

University of Agronomic Sciences and Veterinary Medicine of Bucharest, Faculty of Biotechnology, 59
Mărăști Boulevard, 011464 Bucharest, Romania

Stefania GHEORGHE, Mihai NITA-LAZAR

Control Pollution Department, National Research and Development Institute for Industrial Ecology—
ECOIND, Drumul Podu Dambovitei 57–73, Sector 6, 060652 Bucharest, Romania



For any questions, don't hesitate to contact me via email: laura.feodorov@ecoind.ro